



University
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*POSITIVE INOTROPY IN HUMAN
MYOCARDIUM*

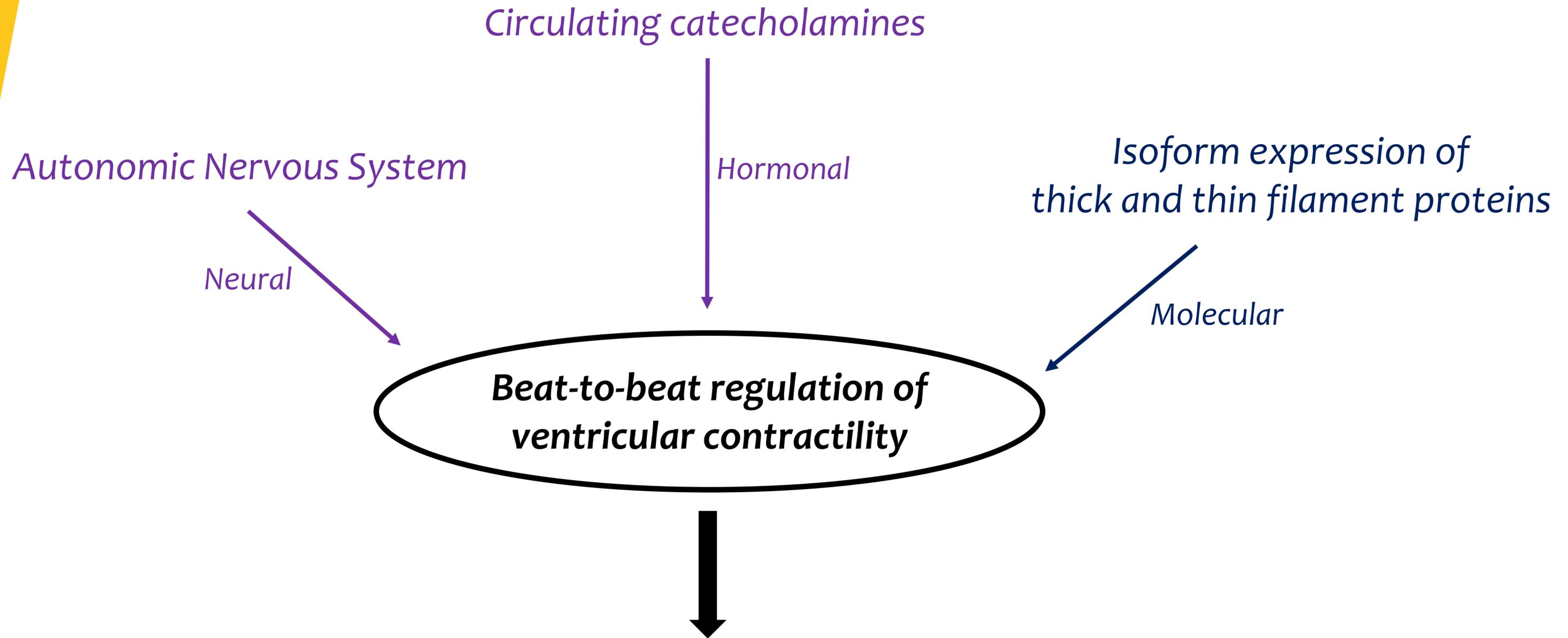
EXPLORING THE ROLE OF MyBP-C

DR. DANIEL FITZSIMONS

ANIMAL, VETERINARY AND FOOD SCIENCES

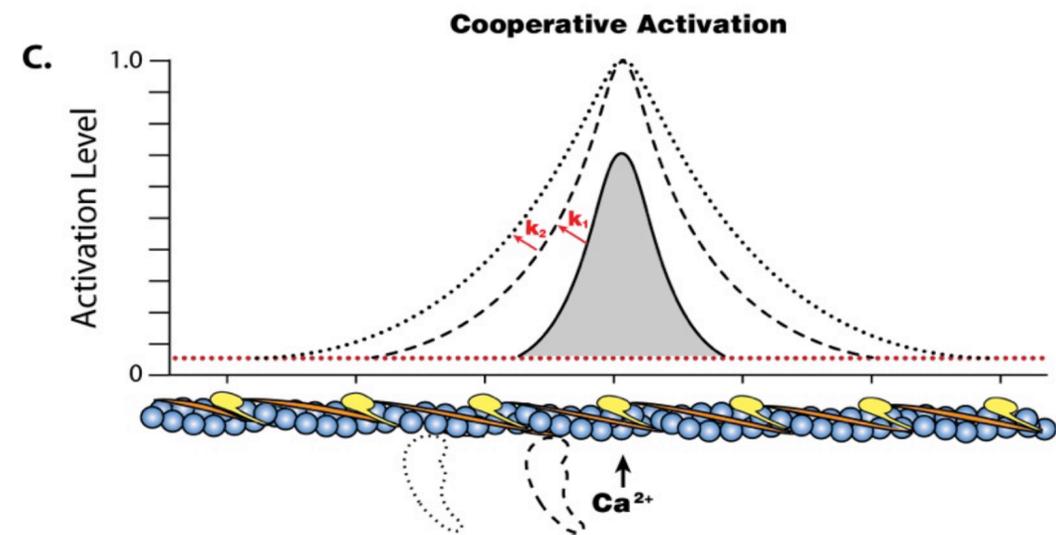
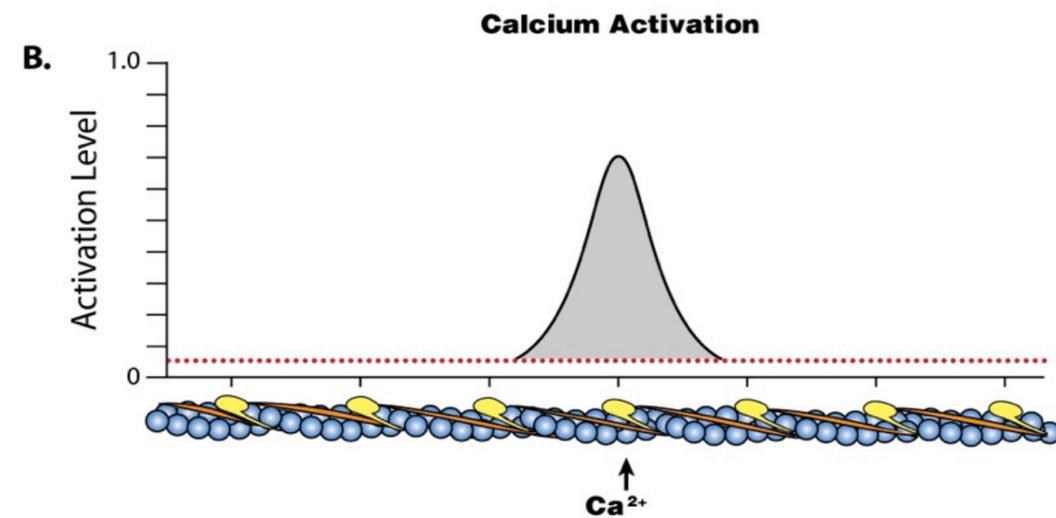
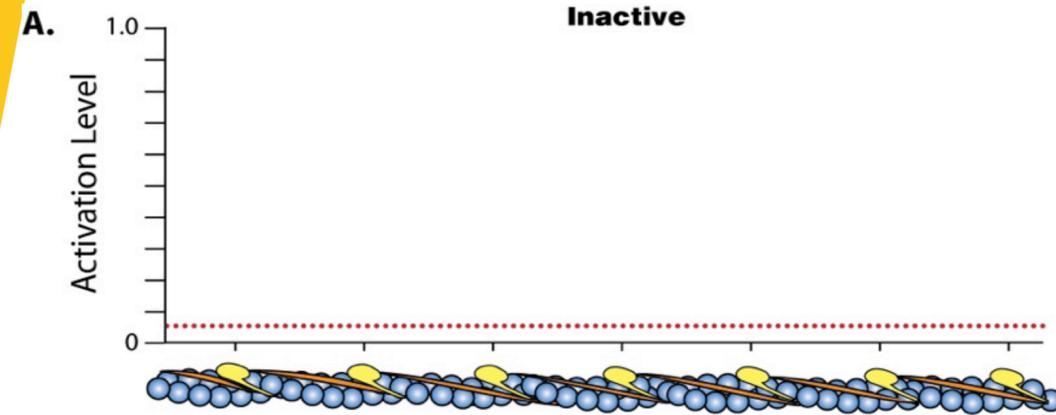
UNIVERSITY OF IDAHO

Ventricular Contractility – A Multidimensional Property of the Heart



Significant variations in the strength of contraction and systolic pressure to ensure that peripheral tissue demands are met.

Diagrammatic Representation of Thin Filament Activation



Initiation of contraction appears to involve the cooperative activation of a short segment of thin filament by Ca^{2+} and / or strong binding myosin cross-bridges, which then activate adjacent regions of thin filament via near-neighbor interactions.

Idea:

Given the nearly ten-fold difference in resting heart rates and myocardial twitch kinetics between rodents and humans, it is highly unlikely that the relative contributions of contractile and regulatory proteins of the thick and thin filament to the cooperative activation of force are the same in these species.

Study:

To test this idea, we examined the Ca^{2+} -dependencies of steady-state force and the rate constant of force redevelopment (k_{tr}) in murine and human ventricular myocardium prior to and following treatment with protein kinase A.

Experimental Workflow

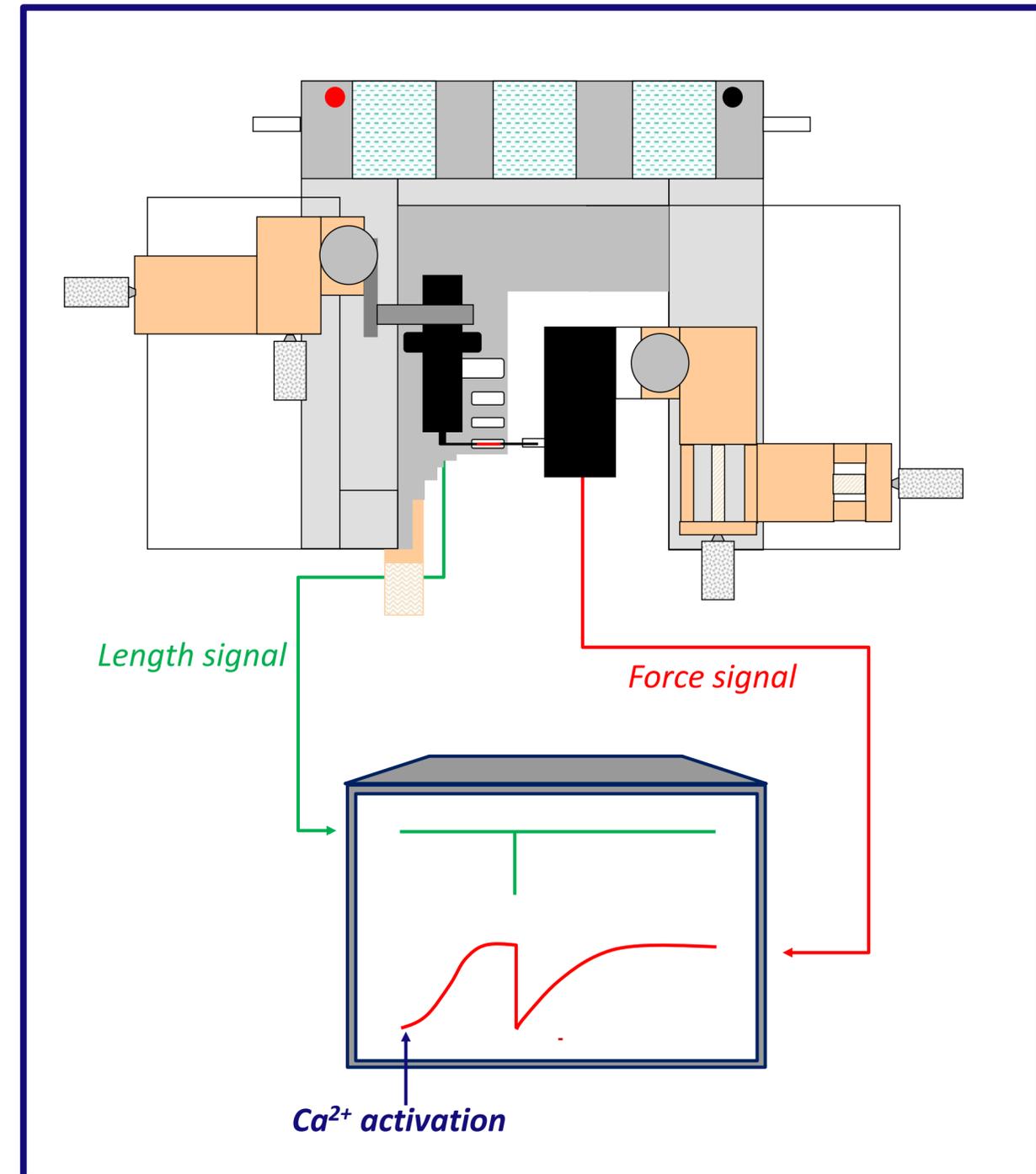


1. Homogenize

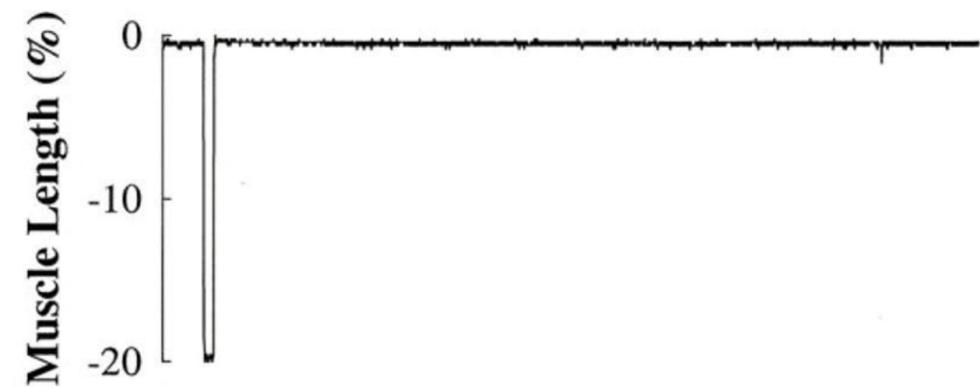
2. Chemically skin

3. Mount

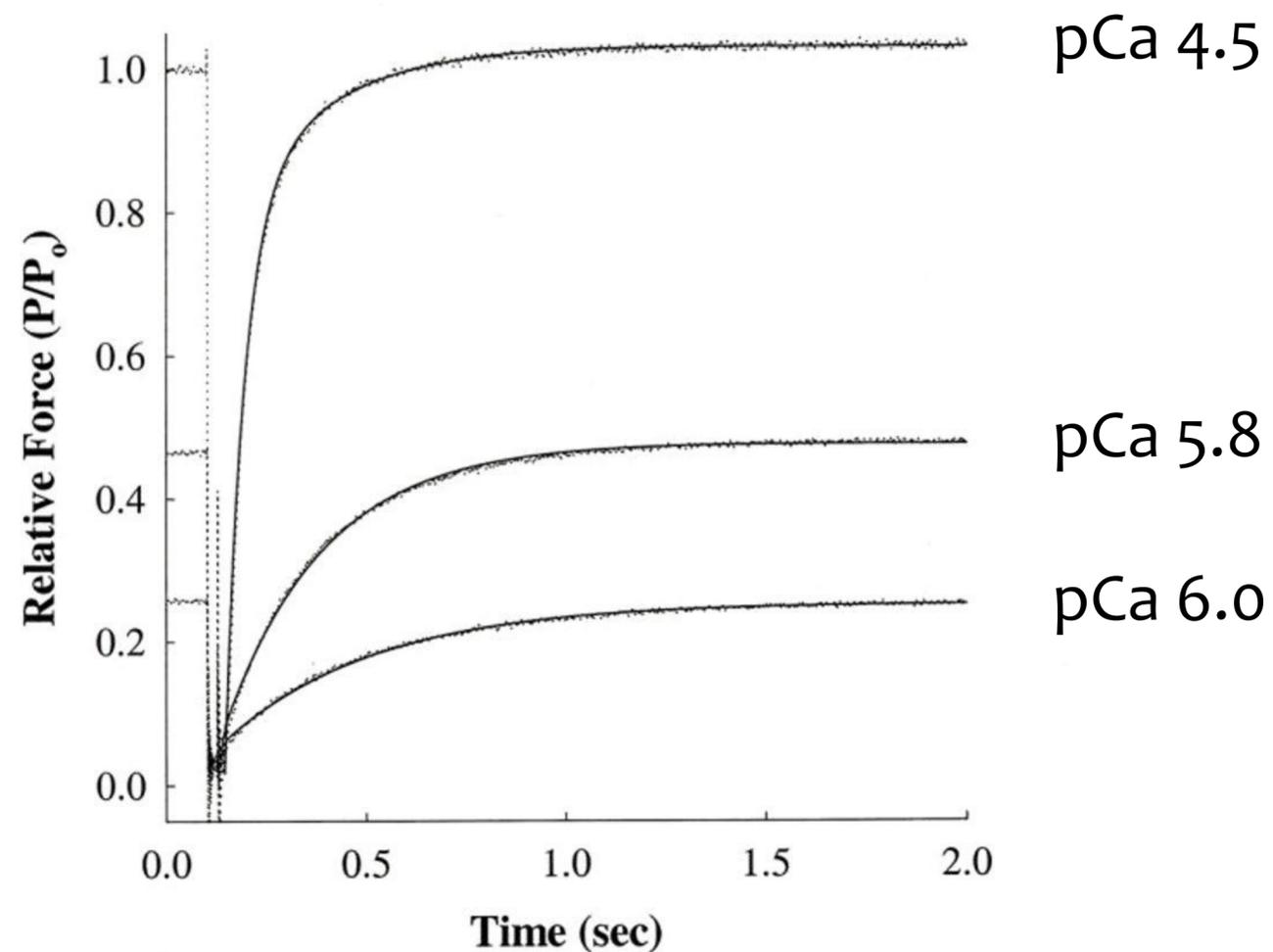
Permeabilized multicellular myocardial preparations
(dimensions: 800-1200 μm x 150-250 μm)



Force Redevelopment in Permeabilized Myocardium



Mechanical release-restretch maneuver to determine the Ca^{2+} -sensitivity of force and the rate constant of force redevelopment at varying levels of Ca^{2+} activation.

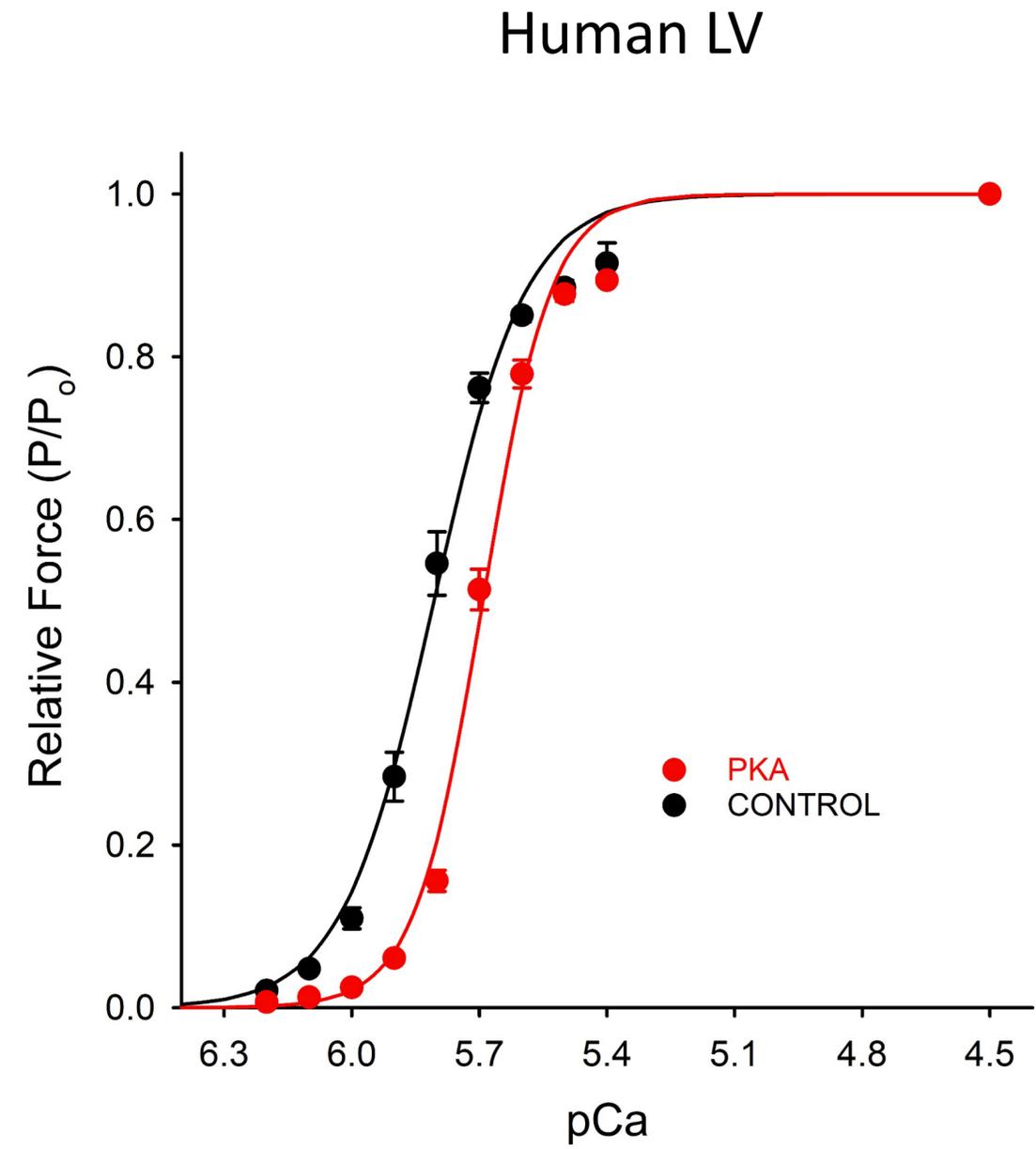
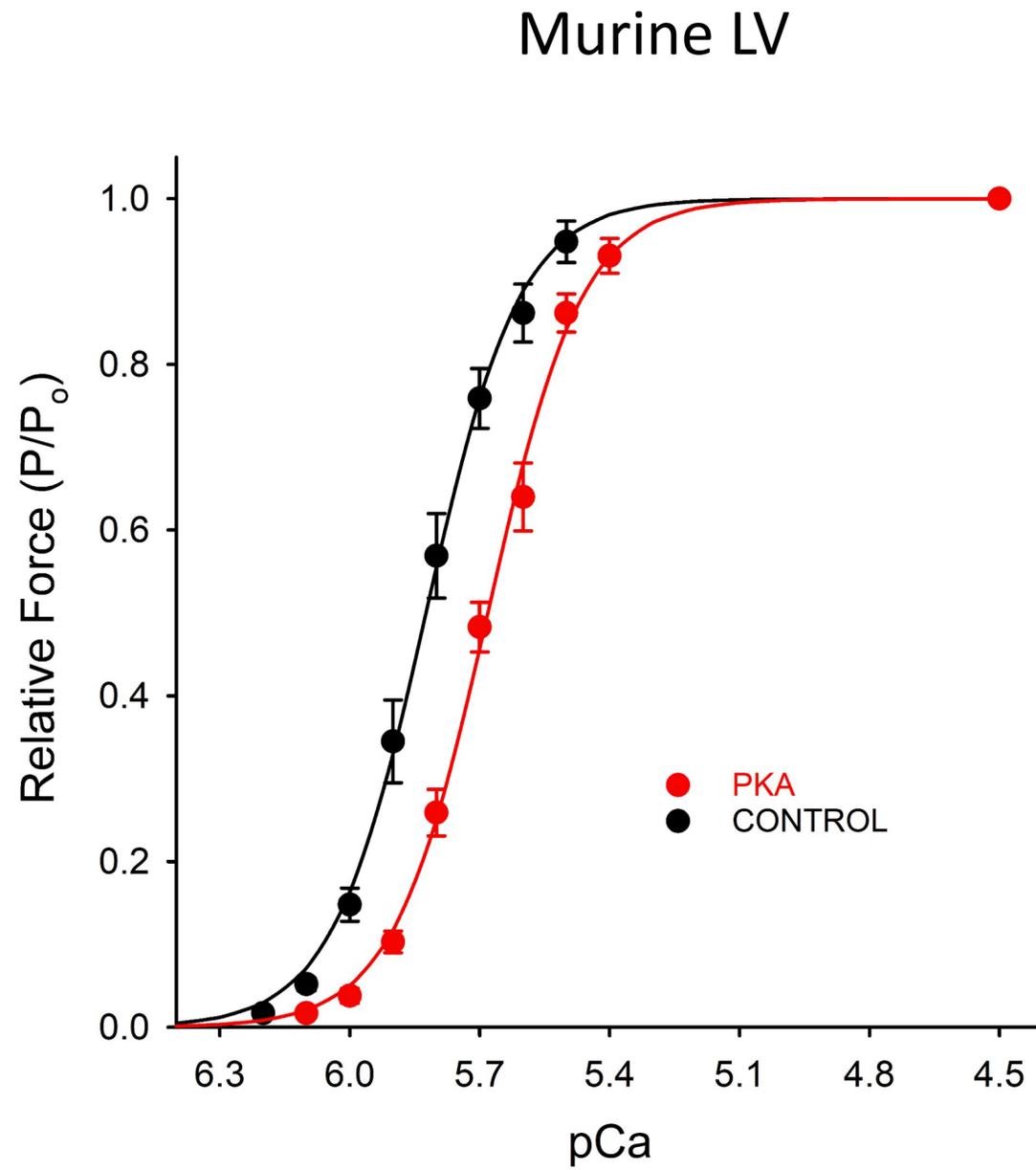


Steady-State Mechanical Measurements



Measurement	Murine LV (Control)	Murine LV (PKA)	Human LV (Control)	Human LV (PKA)
P_o (mN mm ⁻²)	14.4 ± 1.4	12.6 ± 1.4	36.9 ± 2.9	30.5 ± 3.0
P_{rest} (mN mm ⁻²)	0.7 ± 0.1	0.5 ± 0.1	1.8 ± 0.2	1.0 ± 0.2
n_H	4.2 ± 0.2	4.2 ± 0.2	4.3 ± 0.4	5.5 ± 0.1
pCa ₅₀	5.82 ± 0.01	5.69 ± 0.01	5.81 ± 0.01	5.69 ± 0.01
k_{tr} (sec ⁻¹)	26.6 ± 0.9	28.0 ± 1.3	2.7 ± 0.1	2.9 ± 0.2
	└──────────────────────────────────┘ 95% α-MyHC / 5% β-MyHC		└──────────────────────────────────┘ 90% β-MyHC / 10% α-MyHC	

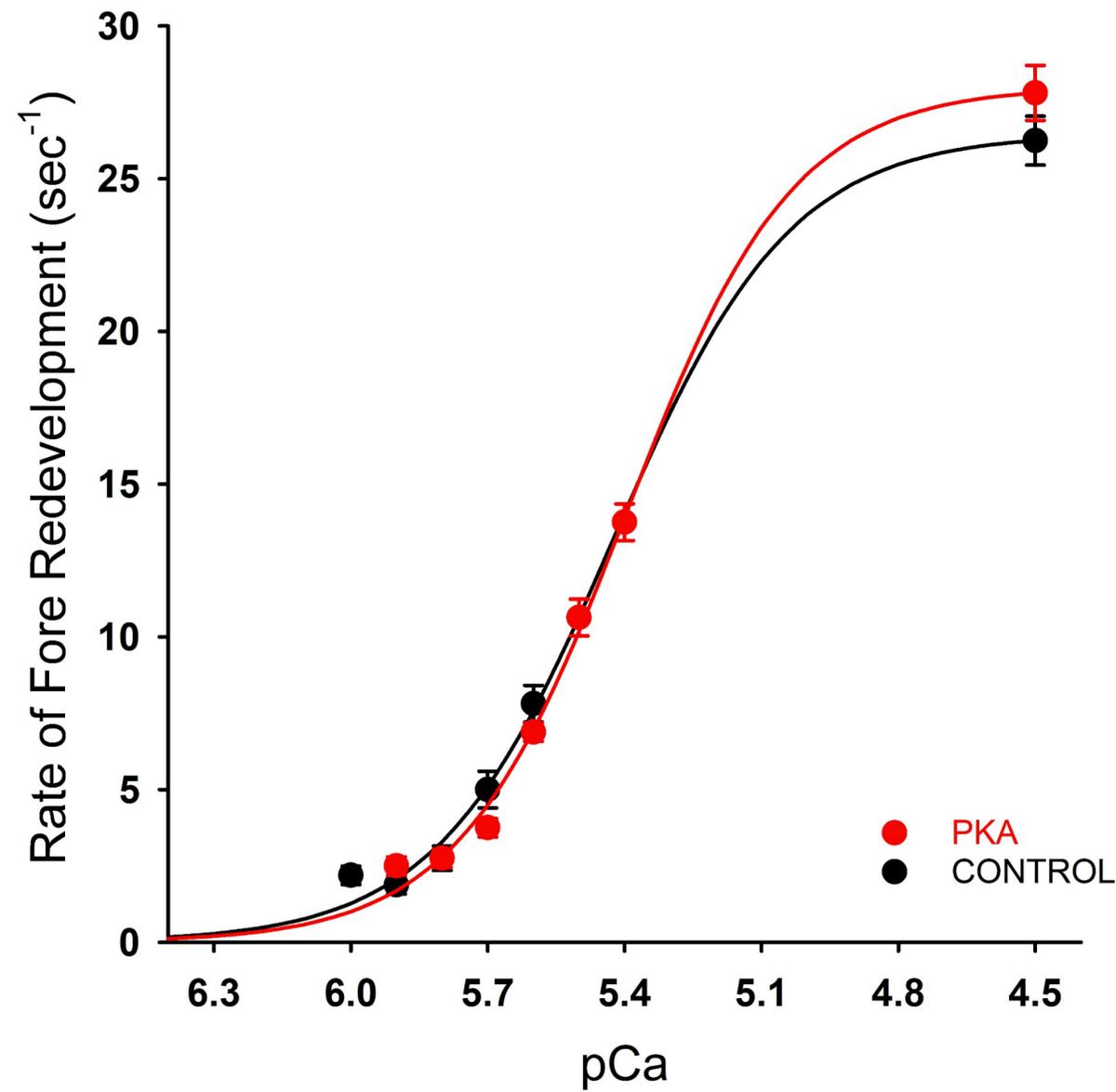
Force-pCa Relationships in Mammalian Myocardium



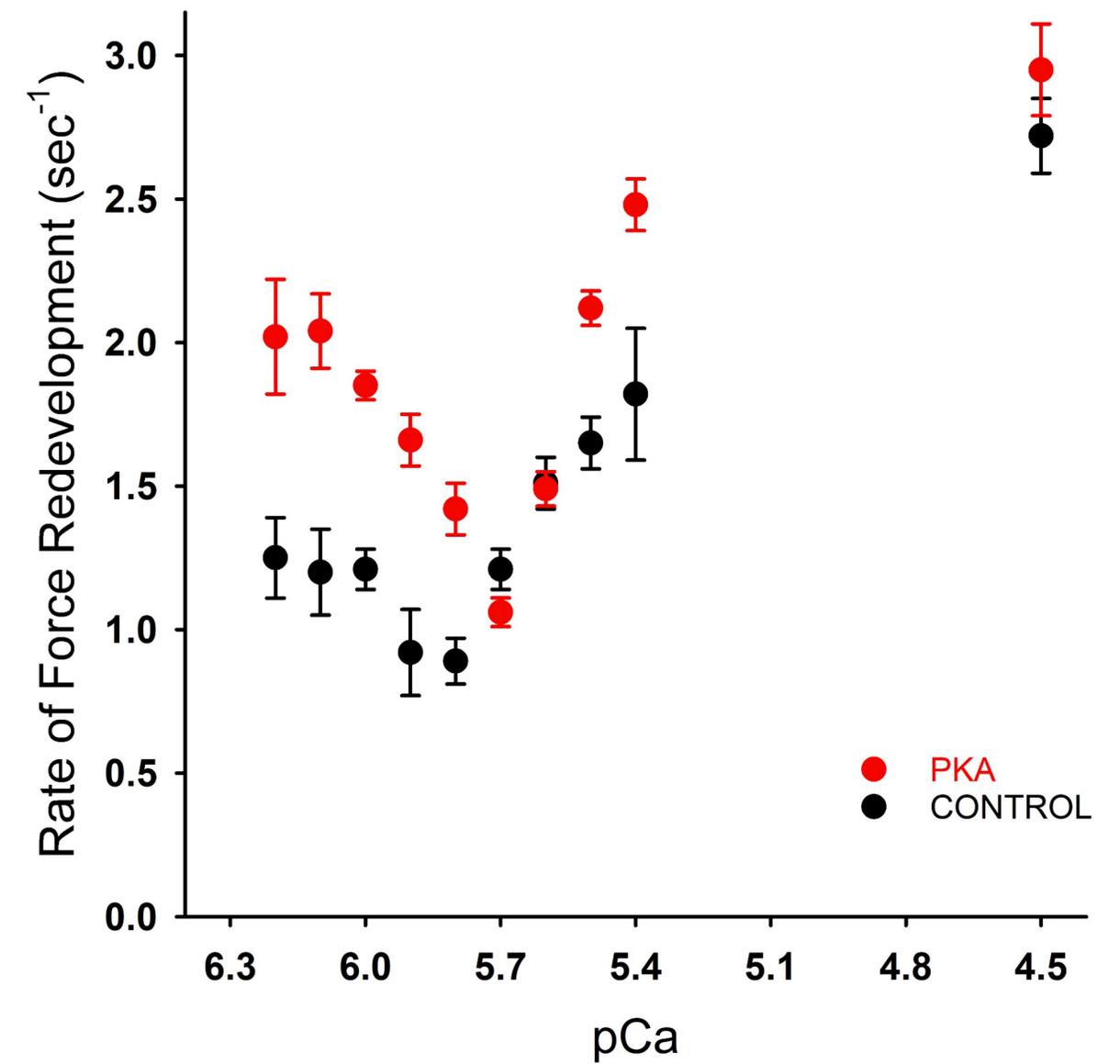
Rate Constant of Force Redevelopment in Mammalian Myocardium



Murine LV



Human LV



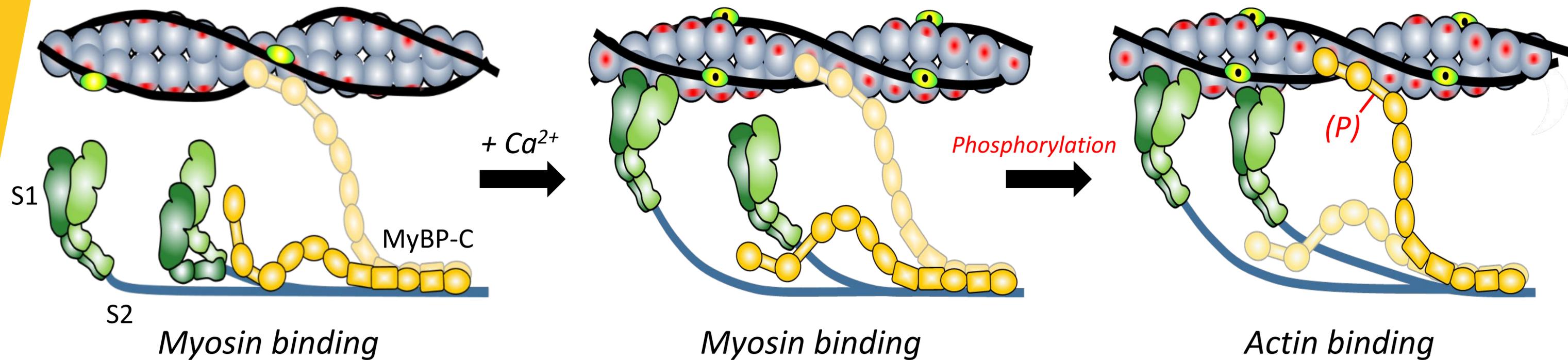
Model for Accelerated Contraction due to MyBP-C Phosphorylation



*Relaxed
Dephosphorylated*

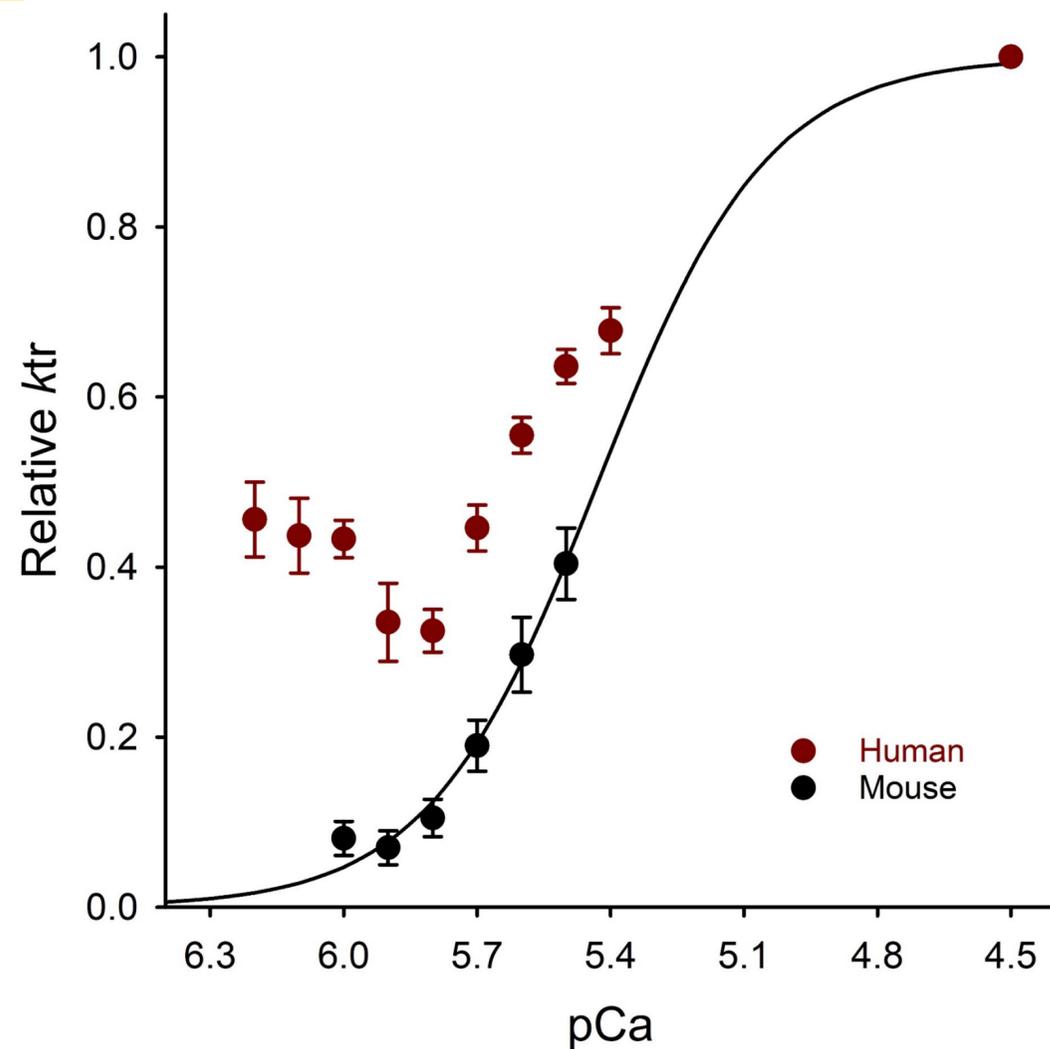
*Ca²⁺-activated
Dephosphorylated
Inhibition of some XBs*

*Ca²⁺-activated
Phosphorylated
Release of XB inhibition*



*Acute modulation of
myocardial work capacity*

Conclusions

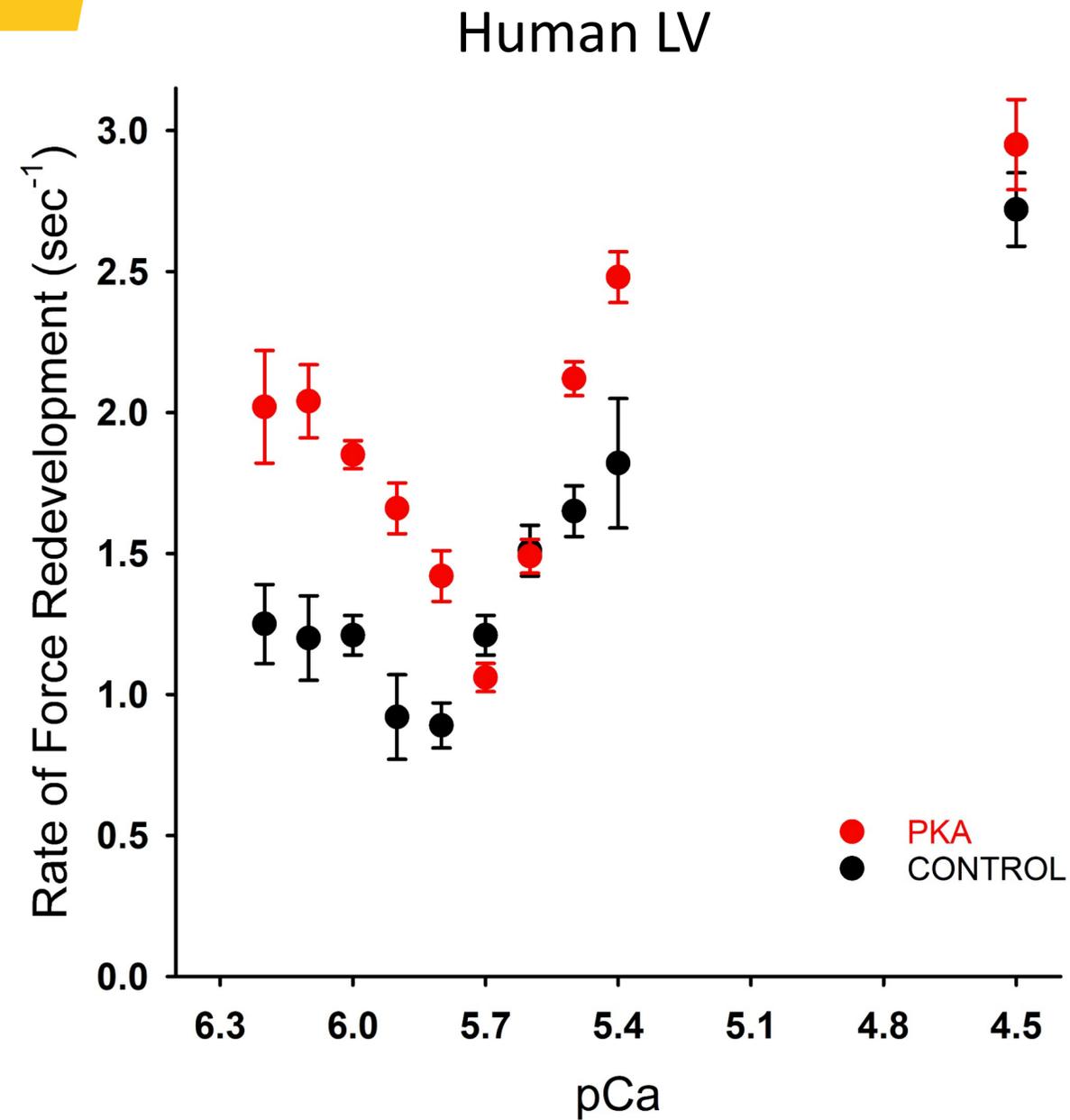


Under control conditions, the faster normalized k_{tr} values at low and intermediate levels of Ca^{2+} activation in human myocardium may reflect an altered level of thin filament activation due to the synergistic expression of α -MyHC and β -MyHC.

When Ca^{2+} is released during EC coupling, α -MyHC is the earliest/first to bind to the thin filament.

Due to its greater activating effect on the thin filament, the binding of α -MyHC opens the thin filament for initial β -MyHC binding and the subsequent cooperative spread of β -MyHC binding.

Conclusions



The effect of MyBP-C phosphorylation on contractile kinetics was amplified in human myocardium.

This may be due, at least in part, to an enhanced level of thin filament activation by the initial binding of fast α -MyHC and phosphorylated MyBP-C to the thin filament.

Thereby, resulting in a greater rate and spread of cooperative β -MyHC binding along the human thin filament.

Acknowledgments



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Thin Filament Regulatory Unit – Myosin Cross-bridge Model (Four-State Model)

